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Effect-based analysis in water/biota

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national research centre for environmental toxicology

Entox is a joint venture between The University of Queensland and Queensland Health

Why use bioanalytical tools for monitoring?

- There are too many chemicals out there to quantify them one-by-one
- In addition: transformation products formed during treatment and in environment
- . Any mixture effects?

Bioassays can be used as sum parameters indicating the overall toxic potential of an unknown chemical cocktail



Schwarzenbach, R.P., Escher, B.I., Fenner, K., Hofstetter, T.B., Johnson, C.A., von Gunten, U. and Wehrli, B. (2006). The challenge of micropollutants in aquatic systems. Science, **313(5790): 1072-1077.**



What is our protection goal?



Gohlke and Portier, Environ Health Perspect 115:1261–1263 (2007)

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There is more to health than cellular effects



BUT: For chemical-induced effects, the initial interaction with the cells is a necessary but not a sufficient precondition



Gohlke and Portier, Environ Health Perspect 115:1261–1263 (2007)

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Conceptual framework: Adverse outcome pathway (AOP)



Adapted from Collin et al. (2008) and Ankley, et al. (2010) in Escher and Leusch, Bioanalytical Tools in Water Quality Assessment, IWA, London, December 2011

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Toxicity pathway



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Bioanalytical tools

- Simulate the toxicokinetics (including metabolism)
- Indicative of the primary interactions with the biological target
 - three main classes of modes of toxic action



- or indicative of adaptive stress response/defense mechanisms
- . Low-complexity or in-vitro bioassays- ideally based on cell lines
- Cost-efficient and high-throughput
 - 96 well plate format
 - reporter gene assays



Bioanalytical test battery

	Mode of action	Assay	Targeted chemicals
Non specific toxicity	Baseline toxicity	Bioluminescence inhibition assay	All chemicals
	General cytotoxicity	Mammalian cell lines, MTS and NRU	All chemicals
Specific	Acetylcholinesterase AChE inhibition	AChE (neurotox)	Organophosphates, carbamates
Receptor	Photosynthesis inhibition	I-PAM (phytotox)	Triazine and phenylurea herbicides
	Estrogenic effects	E-SCREEN	Estrogens, estrogenic industrial chemicals
	Genotoxicity	<i>umuC</i> (genotox)	Aromatic amines, PAH, hard electrophiles (e.g., MMS)
Reactive toxicity	Protein damage	<i>E.coli</i> GSH±	soft electrophiles (e.g., Seanine)
The P	Oxidative stress	Induction of Nrf2 in AREc32	quinones, reactive oxygen species

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What an experiment looks like





relative enrichment factor REF = enrichment factor_{SPE} x dilution factor_{assav}



- Oxley Ck WWTP Inlet
- Oxley Ck WWTP Activated Sludge
- Oxley Ck WWTP post Clarifiers
- Oxley Ck WWTP post UV
- \times Field blank
- + Lab blank







What an experiment looks like





relative enrichment factor REF = enrichment factor_{SPE} x dilution factor_{assav}



- Bundamba AWT Inlet
- Bundamba AWT Post RO
- Bundamba AWT Post AO
- PRW Pipeline
- Mid Brisbane
- Bottled Water
- × Field blank
- + Lab blank





From Sewage to Drinking Water: The Seven Barriers of Water Recycling Barrier 7 Water treatment plant disinfection, distribution and quality management



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Queensland

water Commission

Microtox assay:

bioluminescence inhibition w/ Vibrio fischeri





Non specific

toxicity





Specific (receptor-mediated) toxicity









Genotoxicity – umuC assay







Toxicity reduced across the seven treatment barriers in all bioassays

- Micropollutant burden was reduced by two order of magnitude or more, but to a different extent, in Barriers 2 to 5
- Effects in Barrier 6 and 7 and in drinking water were very low for most endpoints, typically falling below the detection limit or not significantly different from the blank
- Detection limits of the bioassays comparable or lower than the quantification limits of the routine chemical analysis
- Application for
 - benchmarking of different water sources (stormwater, bore water, coal seam gas water)
 - benchmarking of different treatment technologies:



Reungoat, Escher, Macova, Keller (2011). Biofiltration of wastewater treatment plant effluent: Effective removal of pharmaceuticals and personal care products and reduction of toxicity. Water Research, **45(9): 2751-2762.**

Bioanalytical tools for assessing drinking water treatment





Bioanalytical assessment of the formation of DBPs during drinking water treatment

- Full-scale metropolitan drinking water treatment plant
- Nonspecific toxicity and reactive toxicity increased with increase in total absorbable organic halogens (and individual DBPs) during drinking water treatment



Neale, Antony, Bartkow, Farre, Heitz, Kristiana, Tang, Escher, in preparation.

More information: the book "Bioanalytical Tools in Water Quality Assessment"

- Spin-off from industry and regulator's workshops to communicate the scientific basis of bioanalytical tools
- Prepared as part of the development of a risk communication strategy for the Urban Water Security Research Alliance





Escher, B.I. and Leusch, F.D.L., with contributions by CHapman, H and Poulsen, A (2011). Bioanalytical tools in water quality assessment. IWA Publishing, London, UK.

Conclusion

Where we are

- Bioanalytical tools are recognized as valuable research tool
- Bioassays complement
 chemical analysis
- Information on the mixture effects of
 - chemicals
- Wide applicability across the water cycle

The future?

- Evaluate the pollutant burden in biota
- Evaluate the role of transformation products (incl. volatile DBPs?)
- Accepted monitoring tool?
 International harmonisation?
- Bioassay based water quality criteria?