# High Throughput Screening tools for water quality monitoring

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## Introduction

For water quality monitoring, chemical analysis is widely used and well-grounded in regulatory frameworks. However, the practical use of chemical analysis is severely hampered by the need for a priori selection of compounds.

As an alternative, several effect-based bioanalytical tools for water quality evaluation have been developed in FS8.1.1 and 8.1.4.

### Screening results

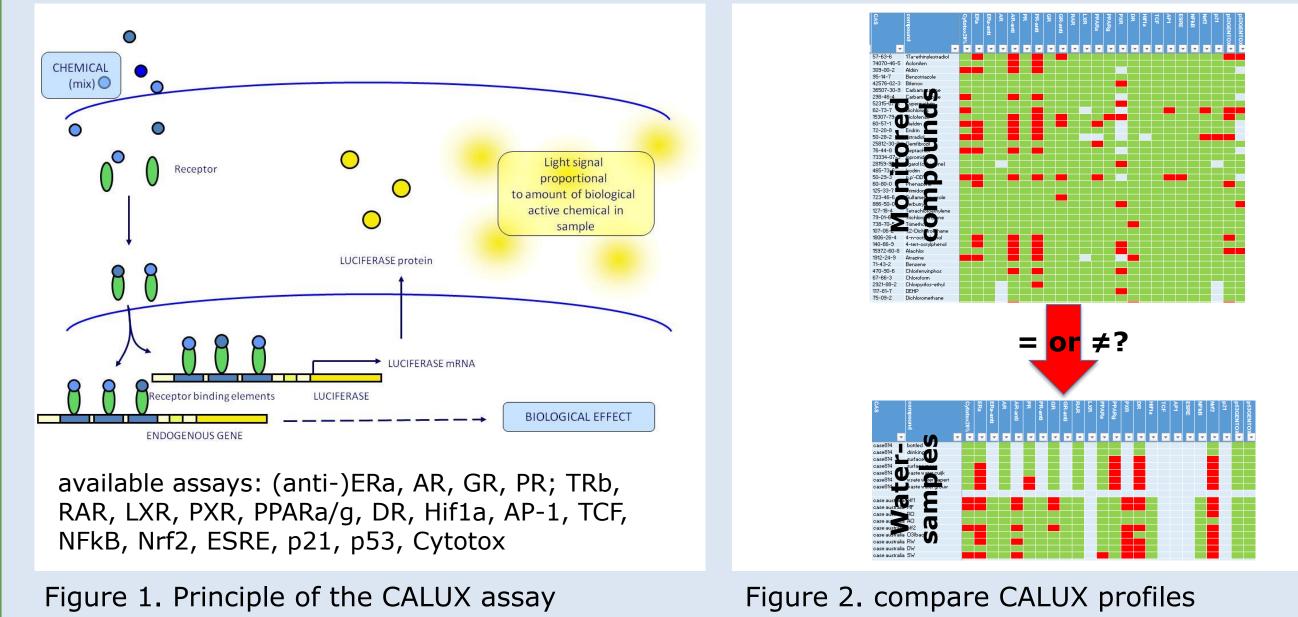
The compounds and the samples activated similar molecular pathways: induction of xenobiotic metabolism, endocrine pathways, and reactive mode-of-action. However, the particular assays activated within these categories were different (table 1). For example, the water samples consistently showed activity on pathways related to oxidative stress (Nrf2) and lipid metabolism (PPAR), while these pathways were hardly activated by the

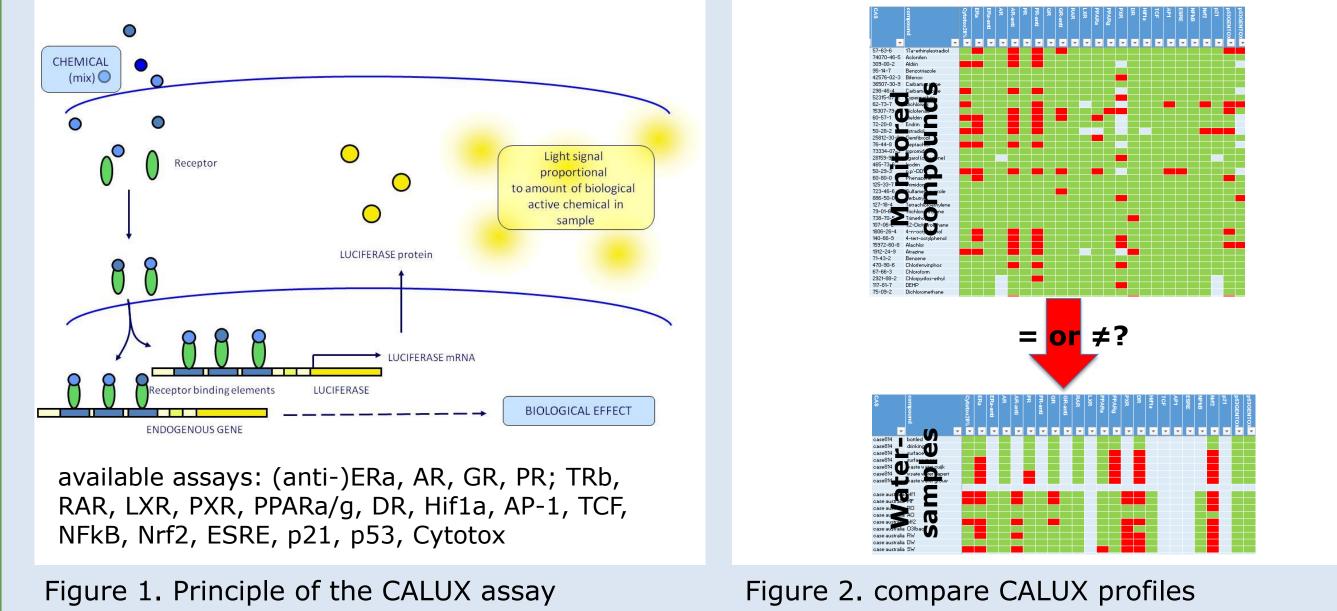
Effect-based assays have the advantage that they provide information on the biological effect of compounds, irrespective of their chemical nature.

## Aim

Recently, two case-studies [1] have been performed where a representative set of water samples was analysed using a broad panel of CALUX reporter gene assays (figure 1).

The aim of the current study was to demonstrate the added value of such an effect-based analysis. To this end, 81 prioritized compounds were screened on the CALUX panel, and the resulting activity profile was compared to the CALUX profile of the water samples (figure 2).





#### routinely monitored compounds.

Molecular pathways	Activated by monitored compounds:	Activated by Water Samples case studies:
Xenobiotic Metabolism:	DR CALUX PXR CALUX	DR CALUX PXR CALUX PPAR CALUX
Endocrine (nuclear hormone receptors):	ERa CALUX antiAR CALUX antiPR CALUX	ERa CALUX antiAR CALUX GR CALUX
Reactive MoA:	p53 CALUX	Nrf2 CALUX
Cell Viability:	Cytotox CALUX	Cytotox CALUX

Table 1. Activated molecular pathways and corresponding CALUX assays for both datasets

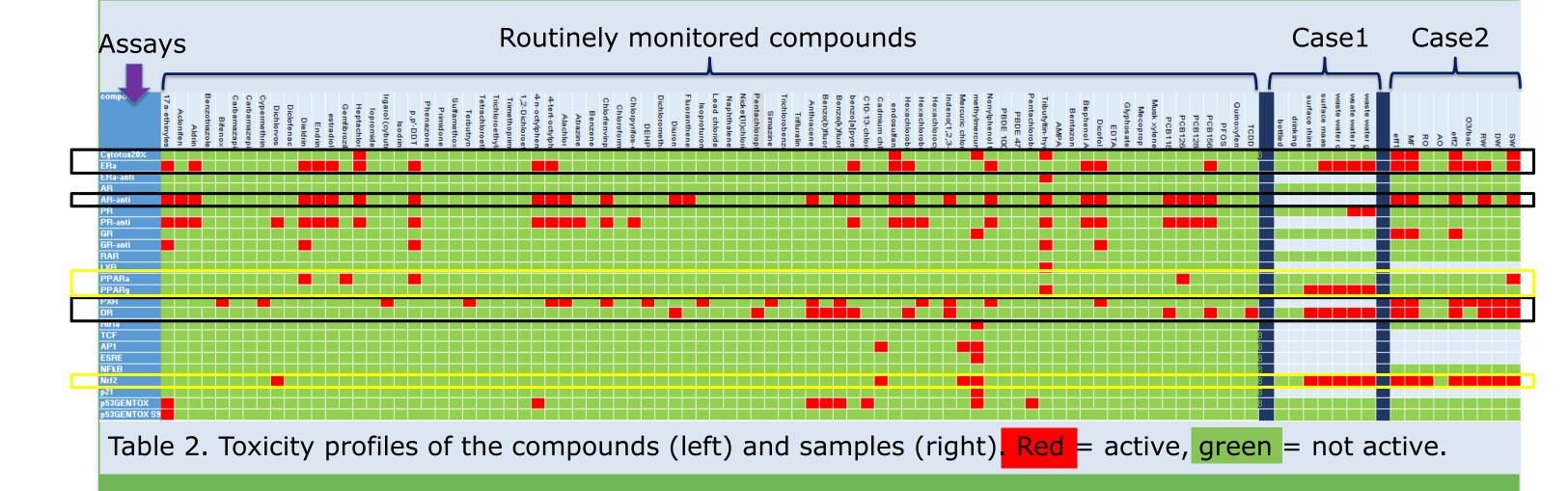
## Activity profiles

Table 2 shows CALUX panel profiles of the compounds (left) and samples (right). The black boxes indicate matches between the two datasets (ERa, antiAR, PXR, DR, Cytotox), while the yellow boxes indicate mismatches (PPARs, Nrf2).



The water samples and a set of 81 routinely monitored substances (figure 3) were measured on the human reporter gene CALUX panel (figure 4), thus generating a toxicity profile.





## Conclusions

The CALUX profile of a set of water samples was compared to the profile of 81 priority pollutants. Although there was considerable overlap between the activated pathways in both datasets, some pathways activated by the water samples were clearly absent in the compounds dataset. This monitored suggests that these compounds cannot fully account for all activities observed in water samples.

The majority of the 81 selected compounds showed activity in at least one CALUX assay. A select panel of CALUX assays could be used to replace laborious chemical analysis; this would enable the detection of the priority pollutants, as well as other compounds that might be present in water samples. The present study clearly shows the added value of effect-based monitoring compared to standard chemical analysis.

	mercuric chloride	15	4-n-octylphenol		PCB156
			4-tert-octylphenol	11	PFOS
1	10 Nonylphenol technical mixture	16	Pentachlorophenol	11	
1	11 Pentachlorobenzene	17	Simazine	12	Quinoxyfen
1	12 PBDE 100	18	Trichlorobenzenes	13	TCDD
	PBDE 47	19	Trichloromethane = chloroform		
1	13 Tributyltin-cation / hydride	20	Trifluralin		

#### Figure 3. Selected compounds: monitored or under review

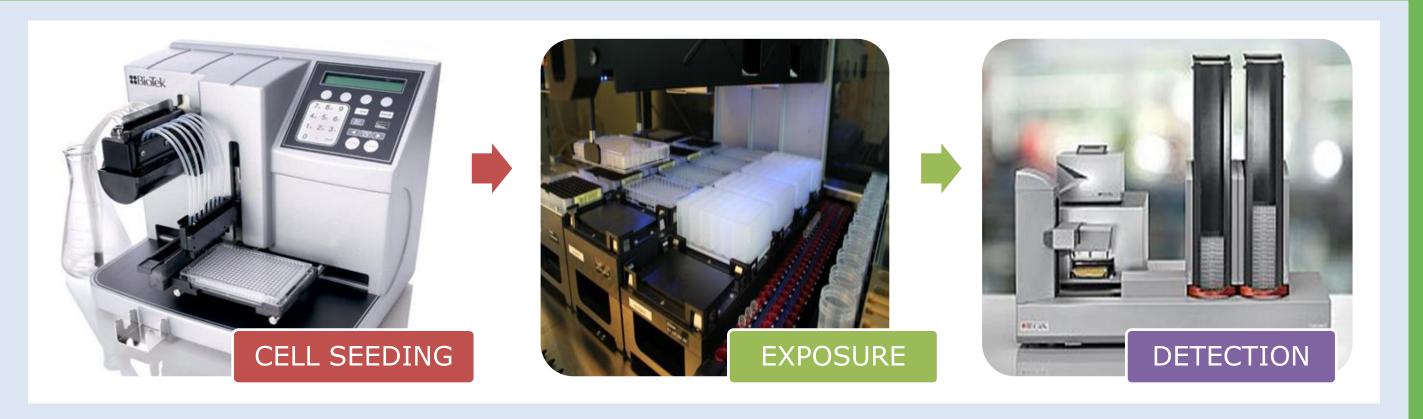


Figure 4. Automated high-throughput screening procedure

## References

[1] Escher BI et al, Environ Sci Technol, 48,1940-56.2014.



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